RESEARCH PAPER

Field assessment of the potentials of some plant-derived insecticide against damage caused by *Leucinodes orbonalis* on eggplant (*Solanum gilo*) at Umudike, Nigeria

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ABSTRACT

The efficacy of plant-derived insecticides was compared with a synthetic insecticide, Lambda-cyhalothrin over two cropping seasons (2015 and 2016) against *Leucinodes orbonalis*, the eggplant fruit and shoot borer (EFSB) infesting eggplant, *Solanum gilo*. The plant materials included oil palm (*Elaeis guineensis*) bunch ash, soursop (*Annona muricata*) seeds, and goat weed (*Ageratum conyzoides*) leaves. These plant materials were formulated into aqueous extracts of 150 g/L (w/v), while Lambda-cyhalothrin was applied at the rate of 5 mL/L (v/v). The experimental design was a Randomized Complete Block Design (RCBD). There were 5 treatments with 3 replications applied on 2 cultivars of eggplants (*Afufa Ukwu* and *Ngwa Large*). Percentage fruit damage varied significantly (P< 0.05) with various treatments. The lowest mean percentage fruit damage was observed in the plot treated with Lambda-cyhalothrin (20.38%), followed by goat weed (29.03%), soursop (40.69%), palm bunch ash (43.30%) and the control (67.08%) for 2015 and 2016. Other parameters measured followed the same trend. Mean yield was significantly affected by the treatments. The highest mean yield was obtained from the plots treated with Lambda-cyhalothrin (12,346.6 g), followed by plots treated with goat weed (11,295.0 g), soursop (9877.2 g), palm bunch ash (8729.4 g) and control was the least (6583.0 g). Among the plant extracts, *A. conyzoides* that recorded low mean percentage damage had very high yield. Therefore, it can be used as insecticides in the control of EFSB.

Key words: eggplant, eggplant fruit and stem borer, insecticides, Leucinodes orbonalis, plant extracts

INTRODUCTION

Eggplant (Solanum gilo) (Family: Solanaceae) commonly called aubergine, brinjal or guinea squash in some parts of the world is an important crop thought to have originated from tropical Africa (PROTA, 2021). Eggplants are popular and important vegetable crop grown year round in most countries of Asia, The Pacific, Europe and Africa (Pugalendhi et al., 2010; Gautam et al., 2019; Nusra et al., 2020). In Nigeria, the crop is identified with many names amongst various ethnic groups; being called Yalo in the North, Igbagba in the Southwest and Anara or Afufa in the Southeast (Ebiringa, 2020). In Southeast Nigeria, immature fresh fruits are eaten raw, and used to welcome visitors into the family (PROTA, 2021). In most other African cultures however, fresh fruit and leaves of the crop serve as vegetables for making soups, porridge and salads (Da-

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vidson & Monulu, 2018). Ethno-medicinally, the roots, leaves and twigs of the plant are used for treating various diseases in most parts of Africa (Han et al., 2021).

One hundred grams of eggplant delivers 25 calories of energy, carbohydrates (6.0 g), protein (1.0 g), fat (0.2 g), sodium (2.0 mg), potassium (229 mg), magnesium (3%), vitamin C (3%) and B-vitamins (5%) (Shukilar & Nalk, 1993; USDA, 2021). Being low in calories (25 Cal/100 g) and fats (0.2 g/100 g), and high in potassium content (229 mg/100 g), studies have shown that eggplant could positively help with reduction of cardiac and liver problems, and to facilitate weight reduction (Oyebade, 2011). Hence, eggplant are suitable for diabetics, hypertensive and obese patients (Prabhu et al., 2009; Pugalendhi et al., 2010, Horna et al., 2007; Gopalan et al., 2007). In some studies in Africa, it was found that eggplant is anticholesterolomic, and protects from glaucoma (Igwe et al., 2003) due in part to certain vitamins and mineral salts thought to help in maintaining the functions of the heart and blood pressure (Oyebade, 2011). Some studies also have associated eggplants with anti-ulcerogenic properties (Chioma et al., 2011). Besides contributing to improved human nutrition, eggplant hold strong potentials to generate income for farmers (Alam

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et al., 2003).

However, the production of this nutrient-rich and healthful vegetable suffers many biotic challenges including diseases and insect pest pressures (Nyeko et al., 2014). Leucinodes orbonalis Guenee (Lepidoptera: Pyralidae) is the most serious and widely distributed insect pest of eggplants (Chakraborti & Sarkar, 2011; Ashadul et al., 2014; Nawaz et al., 2020). It occurs throughout the year and attacks the crop at all stages of its growth (Pugalendhi et al., 2010). Yield losses ranging up to 85-92% have been attributed to attacks of the pest (Patnaik, 2000; Misra, 2008; Jagginavar et al., 2009; Sahu et al., 2020). The larvae could infest eggplants throughout the crop's lifespan (Cork et al., 2001). In the affected crops, the larvae bore into the young shoots, petioles, midribs of large leaves, flowers buds, fruits, and feed on the internal tissues (Singh et al., 2006). The infested flower buds drop and fruits oozed with larval frass through the holes made on the plant, which later becomes portals to secondary fungal infections (Korycinska & Cannon, 2010).

Attempts at curbing this constraint have been done through intercropping with Tagetes sp., crop rotation, use of resistant cultivars (or GM varieties where available), bio-control agents, phytochemicals including pheromones, and frequent sprays of synthetic insecticides (Satpathy & Mishra, 2011; Ashadul et al., 2014; Gautam et al., 2019; Shelton et al., 2019; Goldel et al., 2020). However, intensive applications of insecticide are required for killing the larvae before they bore inside shoots or fruits. Once in the shoots or fruits, EFSB larvae are inaccessible to the killing action of surface-applied chemicals (Alam et al., 2003). Since neonate larvae can enter fruits or shoots within only a few hours of hatching from eggs, pesticides have to be applied frequently in order to have sufficient toxic residues on the plant surface enough to kill the crawling larvae to get effective control effects (Prodhan et al., 2018). Against the backdrop of heavy insecticide application on this crop, indiscriminate use of pesticides, however, has a number of undesirable side effects such as the emergence of resistant species of insects, environmental pollution, and food safety concerns (Goldel et al., 2020; Enyiukwu et al., 2021).

In order to alleviate growing public concerns regarding the effects of synthetic pesticides on environmental and human health, much scientific attention has been geared towards bio-pesticides in recent decades. Bio-pesticides are cheap and environmentally non-disruptive. Therefore, this study was undertaken to determine and compare the effects of plant-derived insecticides (*Annona muricata* seed, *Elais guineensis* bunch, *Ageratum conyzoides* leaves) with a synthetic insecticide (Lambda-cyhalothrin) on damage by *L. orbonalis* on two varieties of *S. gilo* grown at Umudike, Abia State Nigeria.

MATERIALS AND METHODS

Research Site. The study was conducted in the Michael Okpara, University of Agriculture Teaching and Research Farm Umudike. Umudike is located within the tropical rainforest area of the Southeast Agricultural zone of Nigeria and lies within the latitude 05°29'N and longitude 07°32'E with altitude of about 122 m above sea level. The site has an average annual rainfall of 2200 mm and temperature of 23 °C minimum and 32 °C maximum (Agro-meteorological station of National Root Crops Research Institute, Umudike, NRCRI). The trials were laid out during the rainy seasons of the years 2015 and 2016.

Soil Analysis. Analysis of the physical (percentage silt, clay, and sand, texture) and chemical (pH, organic carbon, organic matter, available P, total nitrogen, etc.) characteristics of the Research Farm was carried out at the Soil Science Laboratory of NRCRI, Umudike, using standard protocols as adopted by Ikeogu & Nwofia (2013).

Eggplant Cultivar. Two cultivars of eggplants (*S. gilo*) were used for the study; they are Var. *Afufa* Ukwu obtained from Agwu, Enugu State, and Var. *Ngwa Large* from Ngwa, Abia State, Nigeria.

Field Preparation, Layout and Crop Establishment. The experimental field measuring $28 \times 16 \text{ m}^2$ was slashed and ridged by hoeing during the 2015 and 2016 planting season (April–October). Two week old seedlings of these varieties raised from a nursery nearby were transplanted onto the ridges, and moisture was maintained by a natural rain-fed system. The field was laid out in a randomized complete block design (RCBD) arranged in a split plot with 3 replications. There were 30 plots for the trials, each plot measuring $3.6 \times 3.6 \text{ m}$. The ridges were planted with 24 stands of eggplant/plot with planting spacing pegged at $1.0 \times 0.6 \text{ m}$. The distance between plots and across replicates was maintained at 1 m.

Crop Establishment and Treatment. Treatments formulated and used for the study were oil palm (*E. guineensis*) bunch ash, goat weed (*A. conyzoides*) leaf extract, soursop (*A. muricata*) seed extract at concen-

tration of 150 g/L of water, and a synthetic insecticide (Lambda-cyhalothrin). The procedure for their preparation and concentration are given in Table 1. At 4 weeks after planting (WAP), the seedlings were spray-inoculated at sunset with aqueous formulations of the bio-insecticides and lambda-cyhalothrin; and repeat applications were carried out at flowering.

Data Collection. Data were collected once a week for six weeks in 2015 and 2016 trials on the population of *L. orbonalis* larvae per pod/10 stands, number of holes per pod/10 stands, weight of fruits per plant/plot,

and number of fruits damaged/10 pods/10 stands/ plot which was calculated as:

FD (%) =
$$\frac{n}{N} \times 100\%$$

FD = fruit damage;

n = number of damage fruit;

N = total number of fruit sampled.

Data Analysis. Data obtained were subjected to analysis of variance (ANOVA) using the general linear model (GLM) procedure of SAS version 9 (SAS, 2005).

Table 1. Treatment formulations and rates of application

	**	
Treatments	Formulation	Concentration
Palm bunch ash	150 g of palm bunch ash mixed in a litre of hot water and left for 12 hours and sieved to obtain a uniform solution	150 g/L
Annona muricata	150 g of <i>Annona muricata</i> crushed seed is mixed in a litre of hot water, left for 12 hours and further sieved to obtain an aqueous solution	150 g/L
Ageratum conyzoides	150 g of <i>Ageratum conyzoides</i> crushed leaves was soaked in a litre of hot water and left for 12 hours and sieved to obtain an aqueous solution	150 g/L
Lambda-cyhalothrin E.C.	5 mL	5 mL/L

Table 2. Physico-chemical	properties of the experimental	l site before planting	
			Vaara

Doromotor	Years						
	2015	2016					
Physical properties							
Sand (%)	67.80	64.80					
Silt (%)	11.40	11.80					
Clay (%)	20.80	23.40					
Texture	Sandy clay loam	Sandy loam					
Chemical properties							
рН (Н ₂ О)	5.90	5.80					
Organic carbon (%)	1.02	1.56					
Organic matter (%)	1.76	2.68					
Available phosphorus (cmol/kg)	39.60	68.20					
Total nitrogen (%)	0.09	0.25					
Exchangeable calcium (cmol/kg)	4.00	4.40					
Exchangeable magnesium (cmol/kg)	1.60	1.20					
Exchangeable potassium (cmol/kg)	0.12	0.19					
Exchangeable sodium (cmol/kg)	0.35	0.21					
Exchangeable acidity (cmol/kg)	1.12	1.20					
Exchangeable CEC (cmol/kg)	7.19	7.20					
Base saturation (%)	84.42	83.33					

Separation of significant treatment means was carried out with the least significant difference (LSD) at 0.05 probability level.

RESULTS AND DISCUSSION

Results presented in Table 3 showed the response of the two eggplant cultivars during the 2015 and 2016 cropping seasons to attacks of L. orbonalis. It indicated that the number of larvae/pod recorded on Afufa Ukwu and Ngwa Large were (0.40) and (0.44), respectively in 2015. In 2016, the mean number of larvae/pod in Afufa Ukwu and Ngwa Large was 0.39 apiece. Whereas the values of this variable occasioned by attacks of EFSB on the two varieties were statistically (P< 0.05) not at par in the first year, they were however at par in 2016 (Table 3). The mean number of tunnels per pod due to EFSB attacks recorded on Ngwa Large and Afufa Ukwu were 1.15 and 1.00, respectively in 2015; and 0.97 per pod irrespective of cultivar in 2016. Whereas the number of holes per pod observed on Ngwa Large variety differed significantly from those on Afufa Ukwu, the opposite was statistically (P< 0.05) the case in the 2016 cropping season (Table 3).

The results of the percentage of fruit damaged by EFSB, *L. orbonalis* on the test crop in 2015 and 2016 are presented in Table 3. The results showed that the pest-inflicted fruit profile was generally lower in 2015 than in 2016 for the two varieties. *Ngwa Large* had percentage damage of 38.28% while *Afufa Ukwu* had 39.67%. Similarly, 39.33% and 42.56% fruit damage were recorded respectively on the test cultivars in 2016. Though no significant (P> 0.05) difference was observed between the percentage of fruit damage on *Afufa Ukwu* and *Ngwa Large* due to infestation and activities of *L. orbonalis* in 2015, however, the damage observed on *Afufa Ukwu* was statistically superior to that recorded on *Ngwa Large* in 2016 cropping season.

Leucinodes orbonalis Guenee the eggplant fruit and shoot borer (EFSB) has been rated as a very serious pest of eggplant in eggplant growing regions of The Pacific, Asia and Africa (Ashadul et al., 2014; Prodhan et al., 2018; Nawaz et al., 2020). Screening for and use of resistant (or at least tolerant) varieties is adjudged one of the most important steps to pest management (Chukwu & Enyiukwu, 2021). Findings from this study indicated that *L. orbonalis* successfully attacked and sufficiently damaged both cultivars of eggplant. However, the damage was higher on the Vars. *Afufa Ukwu* than *Ngwa Large*. Data in Table 3, therefore suggest that Var. *Ngwa Large* is comparatively tolerant to infestation and onslaught by EFSB, *L*. orbonalis than Var. Afufa Ukwu.

Genetically influenced biochemical factors are reported to play immense roles in cultivar pest susceptibility or resistance. High contents of certain amino acids, ash, and reducing sugars encourage and promote susceptibility of a cultivar to insect attacks. Conversely, the high presence of certain enzymes such as peroxidases, polyphenol oxidase (PPO), phenyalanine ammonium lyase (PAL), silica, lignin and glyco-alkaloids strongly increase the resistance of a cultivar to pest onslaught (Khorsheduzzaman et al., 2010; Davidson & Monulu, 2018). Therefore the better performance of Var. Ngwa Large over Afufa Ukwu (Table 3) may have been due to higher presence of resistance-promoting compounds in it than in Afufa Ukwu. Findings in this study where Ngwa Large eggplant cultivar out-performed Afufa Ukwu in reducing the level of EFSB infestation and fruit damage is consistent with the report of Kassi et al. (2019) in which eggplant Var. Round Brinjal 86602 showed less susceptibility to infestation and tissue or fruit damage by L. orbonalis than Vars. Round White and Singhnath 656 in a trial.

The results of the mean number of larvae/pod on the eggplant treated with different plant-derived insecticides in 2015 and 2016 are presented in Table 4. The mean number of *L. orbonalis* larvae/pod varied significantly (P< 0.05) among the treatments. The mean number of larvae/pod amongst phyto-pesticides was significantly (P< 0.05) lowest in plots treated with goat weed (0.30), followed by *A. muricata* (0.40), and highest in plots exposed to palm bunch ash which had 0.41; though the later was not statically different from effects of *A. muricata*. On the other hand, Lambda-cyhalothrin had out-performed all the botanical treatments reducing the number of larvae per pod from 0.75 to 0.22. However, all the treatments were superior to the effects observed on the plants in the control experiments.

Regarding the mean number of holes on fruits of the eggplant treated with different plant-derived insecticides in 2015 and 2016, the observed entomotoxic effects varied significantly as presented in Table 4. The number of holes/pod recorded on the treated plants was statistically (P< 0.05) highest in the control (2.06), followed by those treated with Palm bunch ash (1.16), and then *A. muricata* (1.03) and goat weed (0.79) whereas 0.36 was recorded on Lambda-cyhalothrin treated plant in 2015 (Table 4). All the botanical insecticides reduced the number of holes per pod than the effects (2.06) recorded on plants in the control. The results of percentage damage to the eggplant (*S. gilo*) by EFSB, *L. orbonalis* after treatment with plant-derived insecticides in 2015 and 2016 are presented in Table

Table 3. Respo eggpl	nse of eggp ant fruit and	lant varietie stem borer	ss Afufa ukv (EFSB) in	<i>vu</i> and <i>N</i> g 2015 and	wa Large (2016 crop	mean nun ping seasc	nber of lar ons	vae/holes pei	r pod and m	ean perce	entage fruit	c damage) t	o attacks by
		Year				Year				Y	'ear		
Variety	201		2016	Mean	201	5	2016	- Mean	2015		201	9	Mean
	Mean n	umber of la	urvae/pod	-	Mean r	number of	holes/pod	1	Mean pe	rcentage	of fruit dar	nage (%)	
Afufa ukwu	0.4(0.39	0.40	1.0	0	0.97	0.99	39.67		42.5	<u>56</u>	41.12
Ngwa Large	0.44		0.39	0.42	1.1	5	0.97	1.06	38.78		39.3	33	39.06
Mean	0.42		0.39		1.0	8	0.97		39.23		40.5)5	
	$\frac{\mathrm{LSD}}{0.02}^{(0.05)}$	Mean numl	ber of larva	e/pod =	$\frac{\mathrm{LSD}}{0.08}^{(0.05)}$, Mean nu	mber of ho	oles/pod =	$LSD_{(0.05)}$ N	1ean perc	centage dan	nage = 2.0	
	$LSD_{(0.05)}$	Year \times Vari	iety = 0.25		$LSD_{(0.05)}$, Year \times Vi	ariety $= 0$.	14	$LSD_{(0.05)}$ Y	ear × Va	iety = 3.89	-	
Table 4. Effect: Lamb	s (mean nur da-Cyhalotl	nber of larv Irin on eggp	'ae/larvae, f olants natur	ercentage ally infest	truit dam: ed by EFS	age and fr B - Leuciı	uit yield) o nodes orbi	of botanical i nalis in 2015	nsecticides and 2016 c	(palm bu tropping	nch ash, sc seasons	oursop, goa	t weed) and
		Ye	ar		Ye	ar		Yea	r		Ye	ar	
Treatm	lent –	2015	2016	Mean	2015	2016	Mean	2015	2016	Mean	2015	2016	Меан
		Mean nu larvae	tmber of \$/pod		Mean nui holes/	mber of 'pod		Mean perce fruit dama	thage of the of the of the of the of the of the off th		Fruit yi	eld (g)	
Control		0.75	0.63	0.69	2.06	1.67	1.87	70.28	63.89	67.08	4550.0	8616.0	6583.0

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	Mean	(%	41.12	39.06		2.01		goat weed) and		6 Mean		0.06583.0	4.8 8729.4	5.0 9877.2	8.0 11,295.0	7.2 12,346.6	.4	ent = 667.2
ar	2016	f fruit damage ($^{\circ}_{\circ}$	42.56	39.33	40.95	ntage damage =	ty = 3.89	ch ash, soursop, asons	Year	2015 2016	Fruit yield (g)	4550.0 8616	6645.0 10,80	8068.4 11,680	8952.0 13,63	0,106.0 14,58	7666.1 11866	SD (0.05) Treatme
IC	15	percentage of	67	78	23) Mean perce	, Year \times Varie	es (palm bun [.] 6 cropping se		Меан	I J J	67.08	43.3	40.69	29.03	20.28 1		3.18 I
	20	Mean	39.	38.	39.	$LSD_{(0.05)}$	$LSD_{(0.05)}$	l insecticid 15 and 2010	ear	2016	centage o nage (%)	63.89	43.00	40.56	27.50	18.33	38.67	damage =
	Mean		0.99	1.06		oles/pod =	.14	of botanica inalis in 20	Y	2015	Mean per fruit dar	70.28	41.67	40.83	30.56	22.22	41.11	LSD (0.05) %
Γ	2016	of holes/poo	0.97	0.97	0.97	number of h	Variety = 0	fruit yield) cinodes orb		Mean	f	1.87	1.20	0.99	0.70	0.36		umber of
Ica	2015	ean number o	1.00	1.15	1.08	<mark>О _(0.05) Меап r</mark>	O $_{(0.05)}$ Year $ imes$	damage and EFSB - Leu	Year	15 2016	n number of 10les/pod	6 1.67	6 1.25	3 0.96	.09.0 6	6 0.36	8 0.97	(0.05) Mean nu s/pod = 0.13 1.96
	Mean	M	0.40	0.42		OOO = OOO = 0.00	LSI	centage fruit y infested by		Aean 201	Mean 1	0.69 2.0	0.43 1.1	0.41 1.0	0.29 0.7	0.21 0.3	1.0	of LSD holes = 42
	2016	rvae/pod	0.39	0.39	0.39	ber of larvae/	ety = 0.25	ae/larvae, per lants naturall	ar	2016 N	mber of ¹ //pod	0.63	0.44	0.41	0.28	0.19	0.39	Mean number 1 = 0.04 ⁷ ear
rear	5	number of la	0	4	2	, Mean num	Year × Vari	mber of larv. hrin on eggp	Ye	2015	Mean nu larvae	0.75	0.41	0.40	0.30	0.22	0.42	LSD (0.05) larvae/pod LSD Y
	201.	Mean 1	0.4(0.4	0.42	$\underset{0.02}{\text{LSD}}_{(0.05)}$	$LSD_{(0.05)}$	s (mean nur da-Cyhalot)		Jent	112		sh			lothrin		
	Variety		Afufa ukwu	Ngwa Large	Mean			Table 4. Effect Lamb		Treatn		Control	Palm bunch a	Soursop	Goat weed	Lambda-cyha	Mean	

4. Percentage fruit damage varied significantly (P< in which extracts 0.05) amongst the botanical insecticidal treatments. The profile of activity was goat weed extract (30.6%) eggplant by *L. or*

The profile of activity was goat weed extract (30.6%) followed by *A. muricata* extract (40.8%) \geq palm bunch ash (41.7%) and control (70.3%) in 2015. Similarly, Lambda-cyhalothrin recording 22.22% outperformed all the botanical treatments and control in reducing the damage due to the insect pest on the test crop.

This work showed that the botanical insecticides significantly reduced the population, attacks, and damage caused by EFSB, L. orbonalis on the test crop in varying degrees (Table 4). Goat weed demonstrated higher insecticidal activity than the other extracts. Type and concentration of active ingredient(s), and the kind and position of side chains (methyl, hydroxyl, halides etc.) contained on its carbon skeleton affect insecticidal activity of plant extracts (Echeverrigaray et al., 2010; Enviukwu & Awurum, 2013). The concentration of an active ingredient is influenced by solubility in the extracting solvent (Enviukwu et al., 2014; Enviukwu et al., 2016). The higher insect killing potential demonstrated by goat weed in this study may have been due to higher solubility of its active principles in water than those of A. muricata and E. guineensis. It also suggests that the type of active ingredient or its substituents contained in goat weed differed greatly from those in A. muricata and E. guineensis.

However, the marginal difference in insecticidal activity of A. muricata and E. guineensis suggests that compounds contained in these botanicals may have similar structure, substituent or solubility in water; as a result, they may compete for the same binding site or process on the target insect (Enviukwu et al., 2016). The differential toxicity and higher efficacy of Lambda-cyhalothrin (a semi-pyrethroid that acts by elevating sodium channels leading to paralysis and death of affected insects) over goat weed compounds (precocene I and II an anti-juvenile hormone and possible acetylcholenesterase inhibitor) and A. muricata (lipid disrupting acetogenins) may be due to longer persistence on the test crop than goat weed and soursop extracts which are easily degraded by heat and UV-radiation (Dim et al., 2004; Isman & Reffrin, 2014; Lu et al., 2014; Kumar et al., 2018; Hildago et al., 2018; Chahal et al., 2021; Enyiukwu et al., 2021).

Besides directly killing target pests (Ramos et al., 2006), several workers are of the opinion that these natural products prime host tissues to produce phenolic substances that repel or ward-off pest species (Awurum et al., 2016; Nawaz et al., 2020). Findings in this work

in which extracts of *A. conzoides* and *A. muricata* successfully reduced infestation, attacks, and damage on eggplant by *L. orbonalis* is in conformity with reports of Moreira et al. (2007), Kavitha et al. (2008), Mochaiah et al. (2011), and Owusu (2012) who reported ovicidal and insecticidal activities of soursop, papaya and neem leaf and kernel extracts against *L. orbonalis* Guenee in okro and eggplant.

The results of the fruit yield of the test eggplants exposed to the botanical insecticides and Lambda-cyhalothrin in the 2015 and 2016 cropping seasons are presented in Table 4. Whereas Lambda-cyhalothrin had the highest yield of all treatments (10.106 kg), goat weed extract recording (8.982 kg) was next, followed by A. muricata extract (8.068 kg), and lastly palm bunch ash (6.645 kg). The control experiment recording a yield value of 4.550 kg was the least in the 2015 cropping season. The indices of trend of insecticidal activity for all the treatments (botanical and Lambda-cyhalothrin) followed exactly the patterns observed for number of larvae, number of holes and percentage tissue damage per plant observed on the test crop in the 2015 cropping season than in 2016. Generally the pest pressure was higher in 2015 cropping season than 2016 whereas the fruit yield of the crop was lower in the cropping season of the former year than in the latter (Table 4).

Data obtained from this study also showed higher number of larvae, holes and percentage damage of fruit per pod per plant on Afufa Ukwu than Ngwa Large especially in 2016. High ambient temperatures and humidity encourage fecundity of EFSB (Amana & Omoloye, 2012) may be the reason for higher population, attacks and damage by L. orbonalis on the crop in 2015 than in 2016. Certain mineral elements in plant tissues affect its resistance or otherwise to disease and pest attacks (Enviukwu et al., 2021). Phosphorus and calcium are involved in fostering strength, rigidity and development of strong interlocking cross-walls in plant tissues (Okpara, 2014; Amadioha & Nwazuo, 2019) which translates to improved resistance to pests and diseases. Higher available P and Ca in the eggplant rhizospere of 2016, coupled probably with higher ability of Ngwa Large to utilize these growth factors (Han et al., 2021) may explain the better performance of Ngwa Large over Afufa Ukwu. Furthermore the higher reduction of pest pressure and damage by L. orbonalis by the test extracts translated to higher fruit yields of the plant (Table 4), a view also held by other workers (Satpathy & Mishra, 2011; Sahu et al., 2020).

CONCLUSION

This study has revealed that the use of bio-insecticides from *A. muricata* and *A. conyzoides* extracts hold strong and effective potentials for reduction of damage by EFSB and to improve the yield of eggplant. Therefore these plant extracts especially leaf extract of *A. conyzoides* is recommended as a suitable alternative to the synthetic insecticides for the management of *L. orbonalis* and improvement of eggplant productivity in the field.

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AUTHORS' CONTRIBUTIONS

KCE and NFN planned and designed the field experiment. KCE interpreted the data on insect population and plant damage. NFN carried out the experiment in the field, collected relevant data from the field and analyzed them. All the authors played roles in preparing the manuscript. DNE interpreted phytochemical and mode of action aspects of the work and also arranged and edited the manuscript. All the authors have read and approved the manuscript.

COMPETING INTEREST

All authors declare that we have no conflicts of interest related to the publication of this manuscript.

REFERENCES

Alam SN, Rashi MA, Rouf FMA, Jhala RC, Patel JR, Satpathy S, Shivalingaswamy TM, Rai S, Wahundeniya I, Cork A, Ammaranan C, & Talekar NS. 2003. Development of an Integrated Pest Management Strategy for Eggplant Fruit and *Shoot Borer in South Asia*. AVRDC- the World Vegetable Centre, Taiwan.

- Amadioha AC & Nwazuo ED. 2019. Biochemical composition of seed and husk of cowpea (Vigna unguiculata (L.) Walp.) infected by Colletotrichum destructivum O'Gara in storage. Annu. Res. Rev. Biol. 31(1): 1–7. https://doi. org/10.9734/arrb/2019/v31i130034
- Amana O & Omoloye AA. 2012. Planting date of eggplant, *Solanum gilo* and eggplant fruit and shoot borer *Leucinodes orbonalis* infestation. *NJHS*. 17: 14–19.
- Ashadul MI, Hussain MA, Shapla SA, Mehraj H, & Jamal Uddin AFM. 2014. Plant extract for management of brinjal shoot and fruit borer (*Leucinodes orbonalis* Guenee). Am-Euras J. Agric. & Environ. Sci. 14(12): 1409–1414.
- Awurum AN, Enyiukwu DN, & Odoemenam VK. 2016. Influence of plant-gleaned compounds on the initiation and development of fungal diseases of onion (*Allium cepa* L.) in the field. *J. Biol. Agric. Healthc.* 6(9): 71–80.
- Chahal R, Nanda A, Akkol EK, Sobarzo-Sanchez E, Arya A, Kaushik D, Dutt R, Bhardwaj R, Rahman MH, & Mittal V. 2021. Ageratum conyzoides L. and its secondary metabolites in the management of different fungal pathogens. Molecules. 26(10): 2933. https://doi.org/10.3390/ molecules26102933
- Chakraborti S & Sarkar PK. 2011. Management of *Leucinodes orbonalis* Guenee on eggplant during rainy season in India. *J. Plant Prot. Res.* 51(4): 325–328. https://doi.org/10.2478/v10045-011-0053-5
- Chioma A, Obiora A, & Chukwuemeka U. 2011. Does the African garden egg offer protection against experimentally induced ulcer. *Asian Pac. J. Trop. Med.* 4(2): 163–166. https://doi. org/10.1016/S1995-7645(11)60061-8
- Chukwu LA & Enyiukwu DN. 2021. Varietal response and toxicity of aqueous leaf extracts of *Azadirachta indica* to *Phytophtora colocasiae* causing taro leaf blight in Unwana Southeast, Nigeria. *Direct Res. J. Biol. Biotechnol. Sci.* 7: 28–36. https://doi.org/10.26765/DRJBB092186501
- Cork A, Alam SN, Das CS, Ghosh GC, Farman DI, Hall DR, Maslen NR, Vedham K, Phythian SJ, Rouf FMA, & Srinivasan K. 2001. Female

sex pheromone of brinjal fruit and shoot borer, *Leucinodes orbonalis* blend optimization. *J. Chem. Ecol.* 27: 1867–1877. https://doi. org/10.1023/a:1010416927282

- Davidson GI & Monulu AG. 2018. Vitamins and mineral composition of eggplant (*Solanum macrocarpon*) and "Ukazi" (*Gnetum africanum*) leaves as affected by boiling and steaming. *JSRR*. 21(4): 1–8. https://doi.org/10.9734/JSRR/2018/45255
- Dim LA, Funtua II, Oyewale AO, Grass F, Umar IM, Gwozdz R, & Gwarzo U. 2004. Determination of some elements in *Ageratum conyzoides*, a tropical medicinal plant, using instrumental neutron activation analysis. *J. Radioanal. Nucl. Chem.* 261: 225–228. https://doi.org/10.1023/B:-JRNC.0000030962.86191.07
- Ebiringa VDC. 2020. Proximate and micronutrient compositions of four different cultivars of aubergine (*Solanum melongena* L.). *J. Agric. Food Sci.* 18(2): 135–142. https://doi.org/10.4314/jafs.v18i2.10
- Echeverrigaray S, Zacaria J, & Beltrão R. 2010. Nematicidal activity of monoterpenoids against the root knot nematode *Meloidogyne incognita*. *Phytopathology*. 100(2): 199–203. https://doi. org/10.1094/PHYTO-100-2-0199
- Enyiukwu DN & Awurum AN. 2013. Fungitoxic principles and in vitro antifungal activity of extracts from *Carica papaya* and *Piper guineense* on *Colletotrichum destructivum. J. Biol. Sci.* 7: 29 – 36.
- Enyiukwu DN, Amadioha AC, & Ononuju CC. 2021.
 Evaluation of some pesticides of plant origin for control of anthracnose disease (*Colletotrichum destructivum* O'Gara) in cowpea. *Asian J. Agric.* 5(1): 4–11. https://doi.org/10.13057/asianjagric/g050102
- Enyiukwu DN, Ononuju CC, Awurum AN, & Nwaneri JA. 2016. Modes of action of potential phyto-pesticides from tropical plants in plant health management. *IOSR J. Pharm*. 6(7): 1–17. https:// doi.org/10.9790/3013-06710117
- Enyiukwu DN, Awurum AN, & Nwaneri JA. 2014. Efficacy of plant-derived pesticides in the control of myco-induced postharvest rots of tubers and agricultural products: A review. *Net J. Agric. Sci.* 2(1): 30–36.
- Göldel B, Lemic D, & Bažok R. 2020. Alternatives to

synthetic insecticides in the control of Colorado potato beetle (*Leptinotarsa decemlineata* Say) and their environmental benefits. *Agriculture*. 10(12): 611. https://doi.org/10.3390/agricul-ture10120611

- Gopalan C, Sastri BVR, & Balasubramanian SC. 2007. *Nutritive Value of Indian Foods*. National Institute of Nutrition, Indian Council of Medical Research, India.
- Gautam M, Kafle S, Regmi B, Thapa G, & Paudel S. 2019. Management of brinjal fruit and shoot borer (*Leucinodes orbonalis* Guenee) in Nepal. Acta Sci. Agric. 3(9): 188–195. https://doi. org/10.31080/ASAG.2019.03.0632
- Han M, Opoku KN, Bissah NAB, & Su T. 2021. Solanun aethiopicum: The nutrient-rich vegetable crop with great economic, genetic biodiversity and pharmaceutical potential. Horticulturae. 7(6): 126. https://doi.org/10.3390/horticulturae7060126
- Hildago JR, Parellada EA, Bardón A, Vera N, & Neske
 A. 2018. Insecticidal activity of annonaceous acetogenins and their derivatives on *Spodoptera fruiperda* Smith (Lepidoptera: Noctuidae).
 J. Agric. Chem. Environ. 7(3): 105–116. https://doi.org/10.4236/jacen.2018.73010
- Horna D, Smale M, & Falck-Zepeda JB. 2007. Assessing the potential Economic impact of genetically modified crops in Ghana: A methodical framework. IFPRI, Ghana.
- Igwe SA, Akunyili DN, & Ogbogu C. 2003. Effects of *Solanum melongena* (garden egg) on some visual functions of visually active Igbos of Nigeria. J. Ethnopharmacol. 86(2–3): 135–138. https://doi.org/10.1016/s0378-8741(02)00364-1
- Ikeogu UN & Nwofia GE. 2013. Yield parameters and stability of soybean [*Glycine max* (L.) Merril] as influenced by phosphorus fertilizer rates in two utisols. J. Plant Breed. Crop Sci. 5(4): 54–63. https://doi.org/10.5897/JPBCS12.014
- Isman MB & Seffrin R. 2014. Natural Insecticides from the annonacea: A unique example for developing biopesticides. In: Singh D (Ed.). Advances in Plant Biopesticides. pp. 21–33. Springer. New Delhi. https://doi.org/10.1007/978-81-322-2006-0 2
- Jagginavar SB, Sunitha ND, & Biradar AP. 2009. Bioefficacy of Flubendiamide 480 SC against brin-

jal fruit and shoot borer, *Leucinodes orbonalis* Guen. *Karnataka J. Agric. Sci.* 22(3): 712–713.

- Kassi AJ, Javed H, & Mukhtar T. 2019. Screening of different aubergine cultivars against infestation of brinjal fruit and shoot borer (*Leucinodes orbinalis* Guenee). *Pakistan J. Zool.* 51(2): 603–609. https://doi.org/10.17582/journal. pjz/2019.51.2.603.609
- Kavitha VS, Revathi N, & Kingsley S. 2008. Egg susceptibility of brinjal pest, *Leucinodes orbonalis* Geunee to neem extracts. *Asian J. Bio. Sci.* 3(2): 308–310.
- Khorsheduzzaman AKM, Alam MZ, Rahman MM, Mian MAK, & Mian MIH. 2010. Biochemical basis of resistance in eggplant (*Solanum melongena* L.) to *Leucinodes orbinalis* Guenee and their correlation with shoot and fruit infestation. *Bangladesh J. Agric. Res.* 35(1): 149–155. https://doi.org/10.3329/bjar.v35i1.5876
- Korycinska A & Cannon R. 2010. Eggplant Borer, *Leucinodes orbonalis*. Department of Environment, Food, and Rural Affairs, London.
- Kumar B, Misra A, Rawat AKS, Rawat YS, & Srivastava S. 2018. Simultaneous quantification of precocene I and precocene II through highperformance thin layer chromatography validated method in *Ageratum conyzoides* L. germplasms from Western Himalayas. *Phcog. Mag.* 14(Suppl S1): 141–146. https://doi.org/10.4103/ pm.pm_411_17
- Lu XN, Liu XC, Liu QZ, & Liu ZL. 2014. Isolation of insecticidal constituents from the essential oil of Ageratum houstonianum Mill. against Liposcelis bostrychophila Badonnel. J. Chem. 2014: 645687. https://doi.org/10.1155/2014/645687
- Misra HP. 2008. New promising insecticides for the management of brinjal shoot and fruit borer, *Leucinodes orbonalis* Guenee. *Pest Management in Horticulture Ecosystems*. 14(2): 140–147.
- Mochiah MB, Banful B, Fening KN, Amoabeng BW, Offei BK, Ekyem S, Braimah H, & Owusu-Akyaw M. 2011. Botanicals for the management of insect pest in organic vegetable production. Int. J. Entomol. Nematol. 7(4): 001–013.
- Moreira MD, Picanço MC, Barbosa LCA, Guedes RNC, Barros EC, & Campos MR. 2007. Compounds from *Ageratum conyzoides*: Isolation, structural elucidation and insecticidal activity.

Pest Manag. Sci. 63(6): 615–621. https://doi. org/10.1002/ps.1376

- Nawaz A, Gogi MD, Naveed M, Arshad M, Sufyan M, Binyameen M, Islam SU, Waseem M, Ayyub MB, Arif MJ, & Ali H. 2020. In vivo and in vitro assessment of *Trichoderma* species and *Bacillus thuringiensis* integration to mitigate insect pests of brinjal (*Solanum melongena* L.). *Egyptian J. Biol. Pest Control.* 30: 60. https:// doi.org/10.1186/s41938-020-00258-5
- Nyeko P, Stewart J, Franzel S, & Barklund P. 2004. Farmers' experience in the management of pests and diseases of *Calliandra colothyrsus* in Uganda. *UJAS*. 9: 520–529.
- Nusra MSF, Paranagama PA, Amarasinghe LD, & Udukala DN. 2020. Pheromone baited biopesticide for control of *Leucinodes orbonalis* Guenee in brinjal plant. *Front. Biosci. (Elite Ed.).* 12(1): 35–47. https://doi.org/10.2741/e856
- Okpara DA, Muoneke C, Ofor C, Orji R, Ibiam B, Onwuka J, & Ekeleme F. 2014. Eggplant (Solanum sp.) performance in organic and inorganic systems in South-Eastern Nigeria. In: Rahmann G & Aksoy U (Eds.) Proceeding of the 4th ISO-FAR Scientific Conference. 'Building Organic Bridges', at the Organic World Conference. pp. 839–842. ISOFAR, Istanbul, Turkey.
- Owusu DO. 2012. Effect of crude ethanolic leaf extract of soursop, *Annona muricata* L. on eggplant shoot and fruit borer (*Leucinodes orbonalis* Guen.). *Thesis*. School of Graduate Studies, Kwame Nkrumah University of Science and Technology.
- Oyebade W. 2011. *How African Garden Egg Reduces Cholesterol, Poor Sight.* Guardian Newspaper. https://wiyanigerians.blogspot.com/2011/09/ how-african-garden-egg-reduces.html. Access 14 September 2011.
- Patnaik HP. 2000. Flower and fruit infestation by brinjal fruit and shoot borer, *Leucinodes orbonalis* (L.) Guen damage potential vs weather. *Vegetable Science*. 27(1): 82–83.
- Prodhan MZH, Hasan MT, Chowdhury MMI, Alam MS, Rahman ML, Azad AK, Hossain MJ, Naranjo SE, & Shelton AM. 2018. Bt eggplant (Solanum melongena L.) in Bangladesh: Fruit production and control of eggplant fruit and shoot borer (Leucinodes orbonalis Geunee), ef-

fects on non-target arthropods and economic returns. *PLoS ONE*. 13(1): e0205713. https://doi. org/10.1371/journal.pone.0205713

- Prabhu M, Natarajan S, Veeraragavathatham D, & Pugalendhi L. 2009. The biochemical basis of shoot and fruit borer resistance in inter-specific progenies of brinjal (*Solanum melongena*). *Eur-Asia J. BioSci.* 3: 50–57.
- PROTA (Plant Resources of Tropical Africa) 2021. Solanum aethiopicum. https://uses.plantnet-project.org/en/Solanum_aethiopicum_(PROTA). Accessed 4 July 2021.
- Pugalendhi L, Veeraragavathatham D, Natarjan S, & Praneetha S. 2010. Utilizing wild relative (Solanum viarum) as resistant source to shoot and fruit borer in brinjal (Solanum melongena Linn.). Electron. J. Plant Breed. 1(4): 643–648.
- Ramos MV, Banderia GdP, de Freitas CDT, Nogueria NAP, Alencar NMN, de Sousa PAS, & Carvalho AFU. 2006. Latex constituents from *Calotropis* procera (R.Br.) display toxicity upon egg hatching and larvae of *Aedes aegypti* (Linn). *Mem. Inst.Oswaldo Cruz.* 101(5): 503–510. https:// doi.org/10.1590/S0074-02762006000500004
- Sahu AK, Padhy D, Ramalaxmi V, & Dash L. 2020. An overview of bio-rational approaches for brinjal insect pest management. *Biosc. Biotech. Res. Comm.* Special Issue. 13(12): 89–91.
- SAS Institute. 2005. Statistical Analytical System SAS/STAT User's Guide Version 8(2) Cary NC,

SAS Institute Inc.

- Satpathy S & Mishra DS. 2011. Use of intercrops and antifeedants for management of eggplants shoot and fruit borer, *Leucinodes orbinalis* (Lepidoptera: Pyralidae). *Int. J. Trop. Insect Sci.* 31(1–2): 52–58. https://doi.org/10.1017/ S1742758411000154
- Shelton AM, Hossain MdJ, Paranjape V, Prodhan MdZH, Azad AK, Majumder R, Sarwer SH, & Hossain MdA. 2019. Bt brinjal in Bangladesh: the first genetically engineered food crop in a developing country. *Cold Spring Harb Perspect. Biol.* 11(10): 13. https://doi.org/10.1101/cshperspect.a034678
- Shukilar V & Nalk LB. 1993. Agro-techniques of Solanaceous vegetables. In: Chadla KI & Kallio G (Eds.). Advances in Horticulture Vegetable Crops part I. pp. 365–399. Malhotra Publication House.
- Singh RK, Mittal PK, & Dhiman RC. 2006. Laboratory study on larvicidal properties of leaf extract of *Calotropis procena* (Family-Asclepiadaceae) against mosquito larvae. J. Commun. Dis. 37(2): 109–113.
- USDA (United States Department of Agriculture) 2021. Eggplant, raw. https://fdc.nal.usda.gov/ fdc-app.html#/food-details/169228/nutrients. Accessed 14 July 2021.